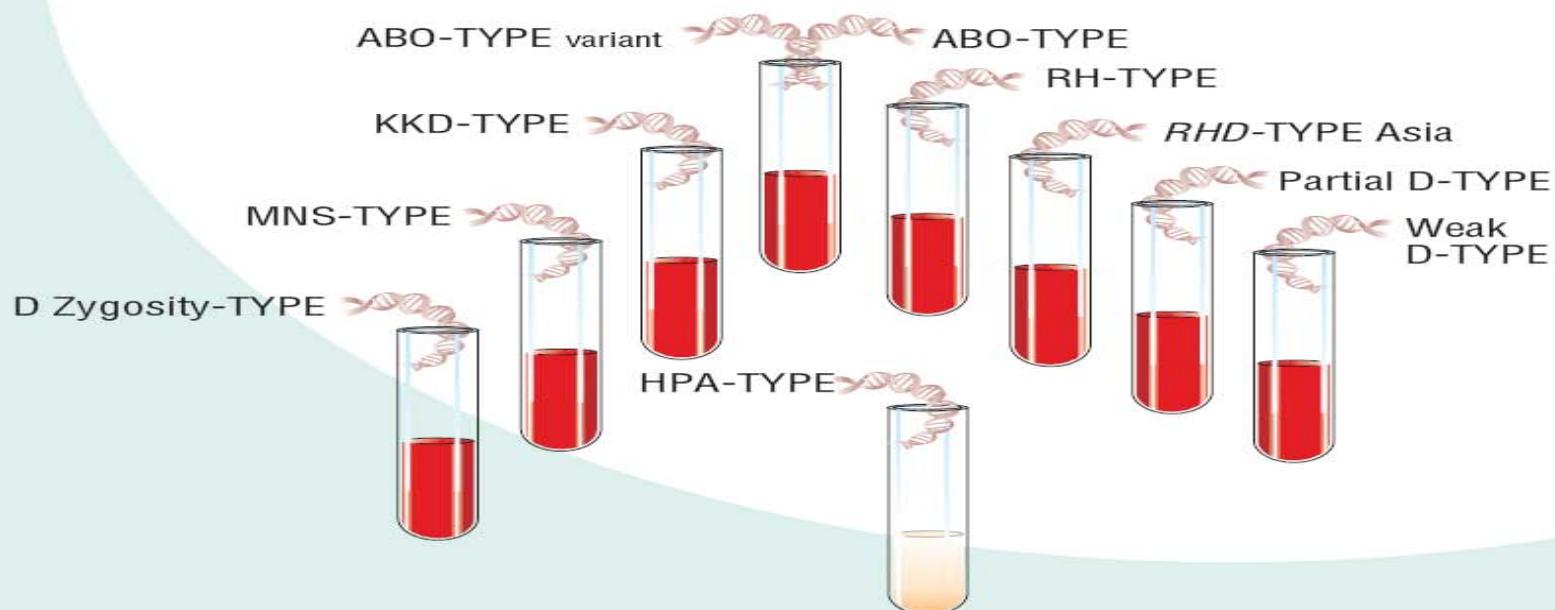


BAGene

SSP kits for determination of

- **ABO blood groups**
- **RH types**
- **Kell, Kidd, Duffy systems**
- **MNS system**
- **HPA specificities**

on a molecular genetic basis

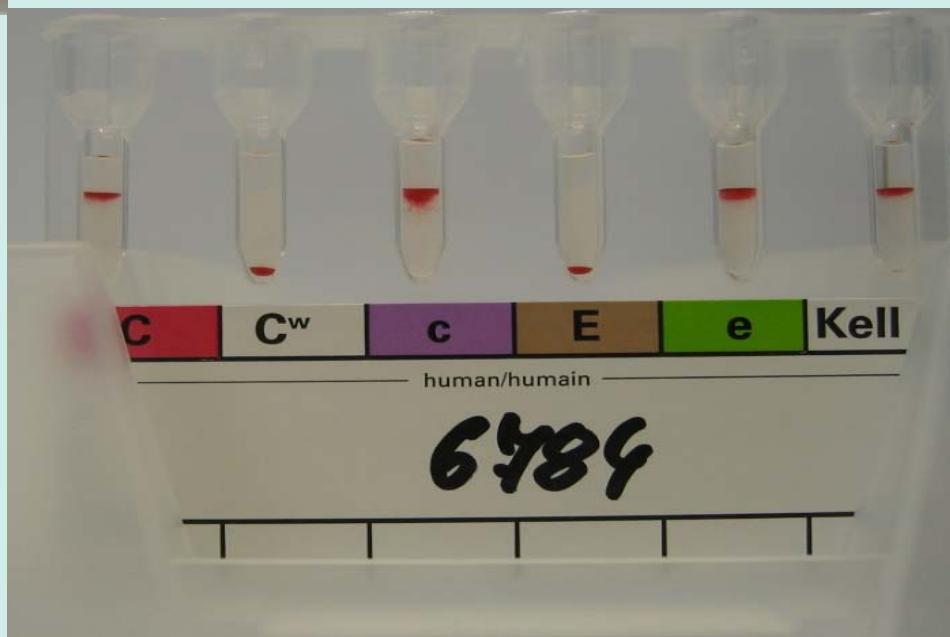
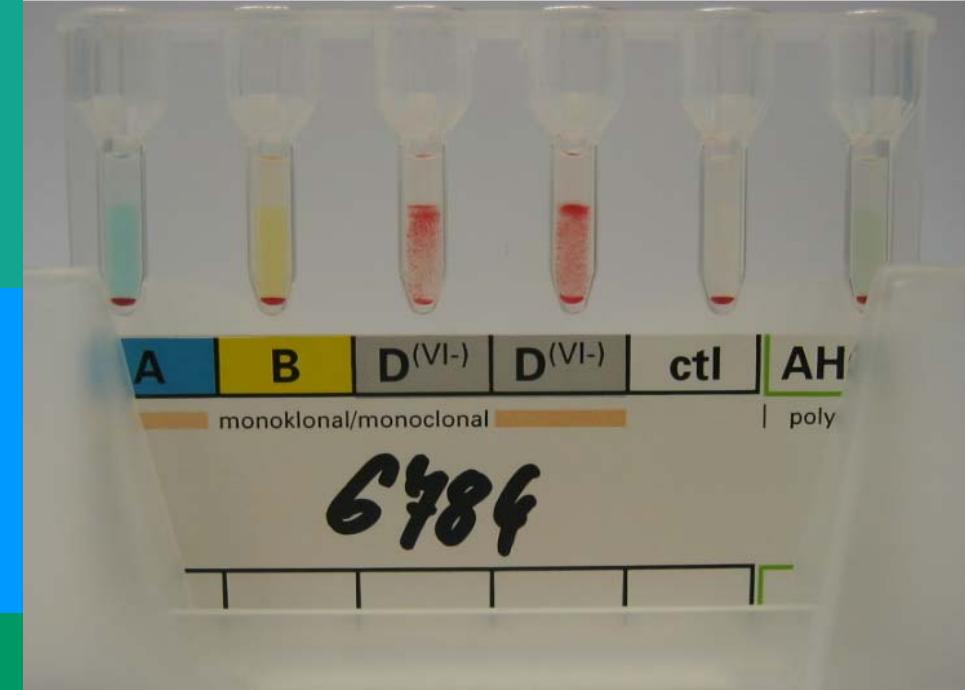


Application

- ♥ Genotype multi-transfused recipients
- ♥ Genotype patients after ABO-incompatible bone marrow transplantation
- ♥ Determine *RHD* zygosity of partners of alloimmunized D-negative women before pregnancies
- ♥ Genotype Rh D negative donors with C or E in order to exclude the presence of the *RHD* gene and thus preventing anti-D alloimmunization of recipients caused by hidden Rh D variants in RBC units
- ♥ Identify genotype in case of weakly expressed Rh D (e.g. DEL) in donors
- ♥ Confirm weak D genotype in recipients in order to avoid the donation of Rh D negative blood units
- ♥ Quality control of serological methods
- ♥ External Quality Assurance trials

Result according to
Transfusion guideline

Rhesus D positive
CcD.ee Kell positive



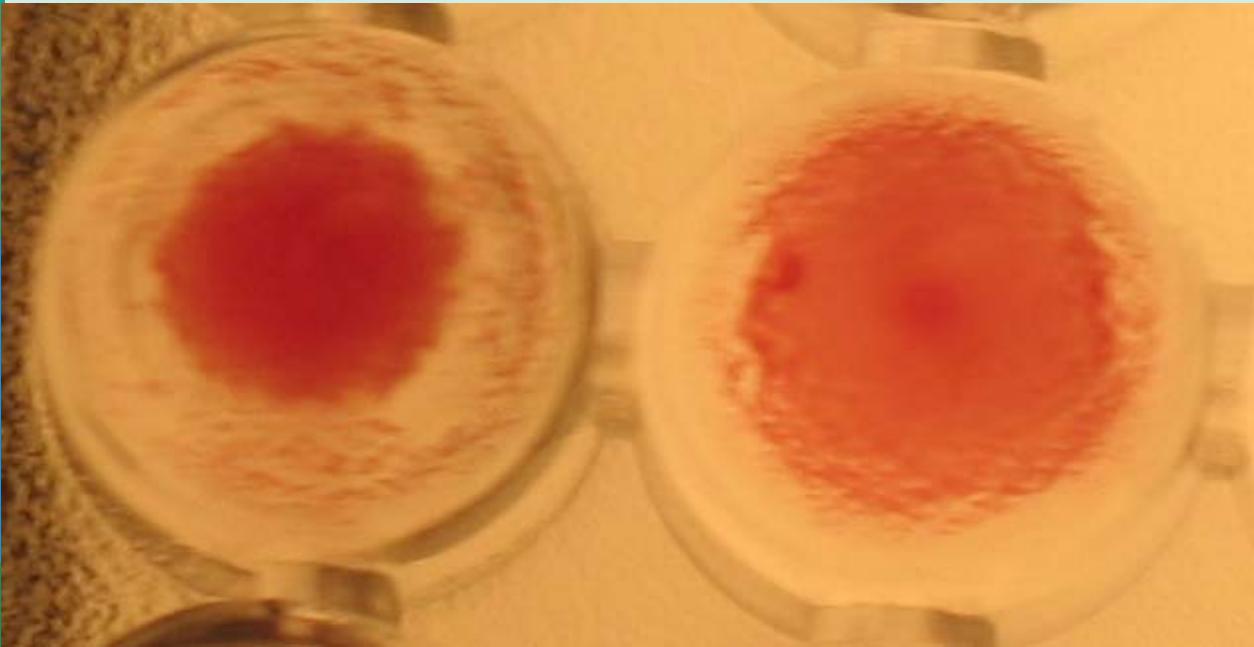
C c D E e ctl

human/humain

E000 206 10.95

M

9031 - 6784



micowell plate
antisera
dilution 1:5

agglutination
strength (1) -1



tube test
immediate spin

questionable
agglutination
with monoclonal
Anti-D

1031-5774

Weak D-TYPE

Worksheet und Auswertetabelle / Worksheet and Evaluation diagram

Reaktions-Nr. / Reaction No.	1	2	3	4	5	6	7	8
PCR-Produkt (Größe in bp) PCR product (size in bp)	150	126	165	101	130	112	198	153
weak D Allele / weak D alleles								
weak D type 1	+	-	-	-	-	-	-	-
weak D type 2	-	+	-	-	-	-	-	-
weak D type 3	-	-	+	-	-	-	-	-
weak D type 4.0, 4.1	-	-	-	+	-	-	-	-
weak D type 4.2, DAR	-	-	-	+	+	-	-	-
weak D type 5	-	-	-	-	-	+	-	-
weak D type 11 (haplotyp cDe)	-	-	-	-	-	-	+	-
RHD(M295I) (haplotyp CD _e e)	-	-	-	-	-	-	+	-
weak D type 15	-	-	-	-	-	-	-	+
RHD pos. oder / or RHD neg.	-	-	-	-	-	-	-	-

Ergebnis Result	1	2	3	4	5	6	7	8	Genotyp Genotype

Proben-ID / Sample-ID: 11 9031 6784

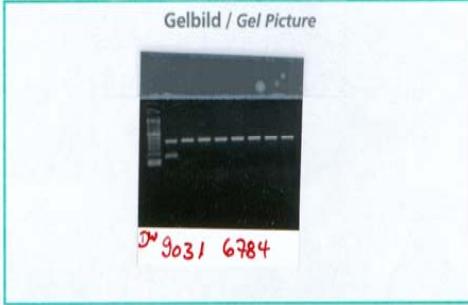
Name:

Geb.-Datum / Birthdate:

Ergebnis / Result: DW typ 1

Datum / Date: 22.10.04

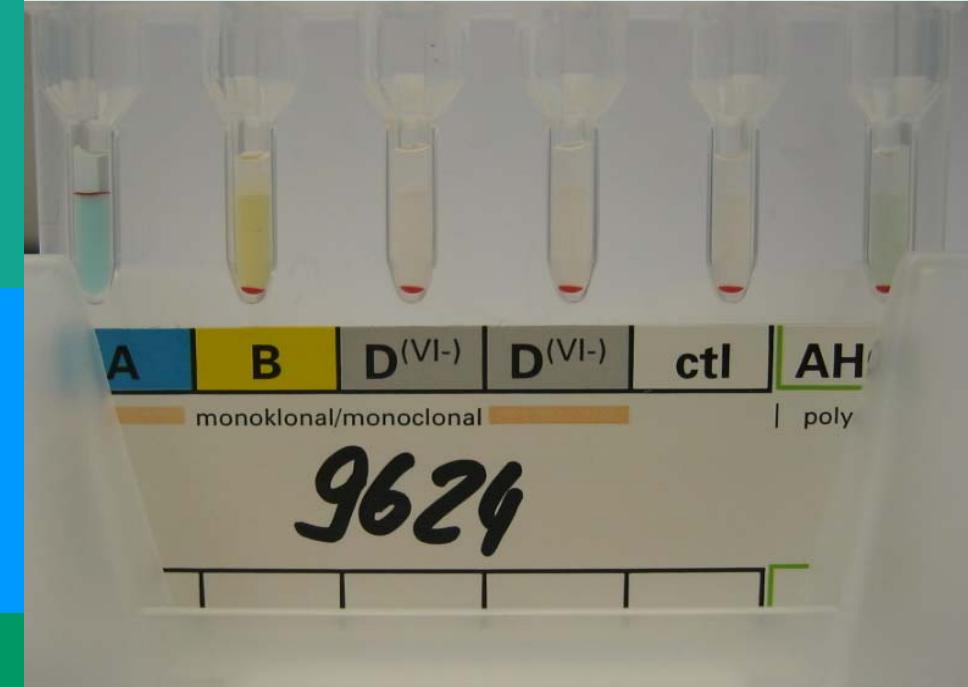
Unterschrift / Signature: Wämmer P.



Final result
weak D type 1

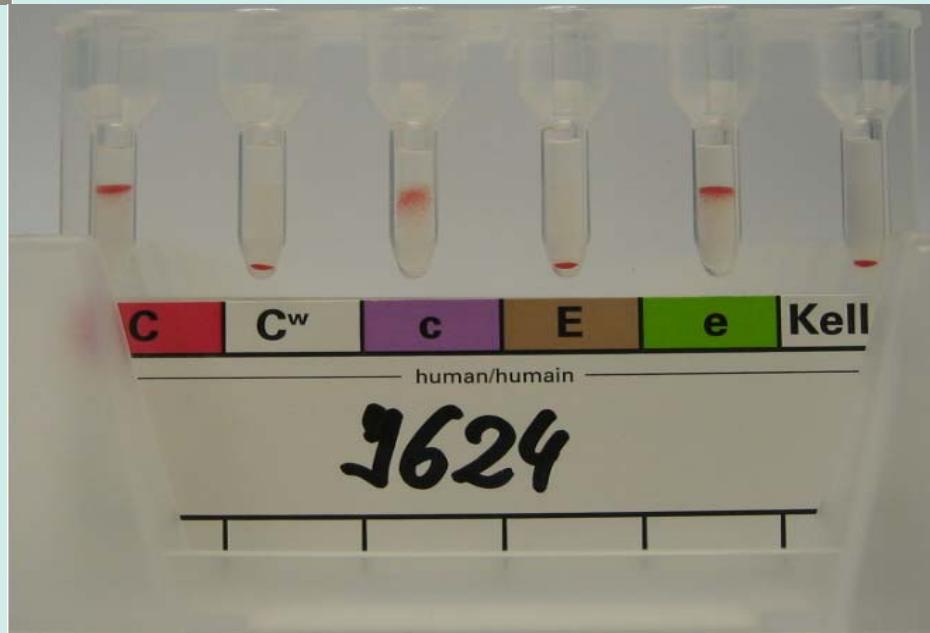
Rhesus prophylaxis
not required

Transfusion of D
positive red blood
cells is possible



Result according to
Transfusion guideline

Rhesus D negative,
Ccdee Kell negative



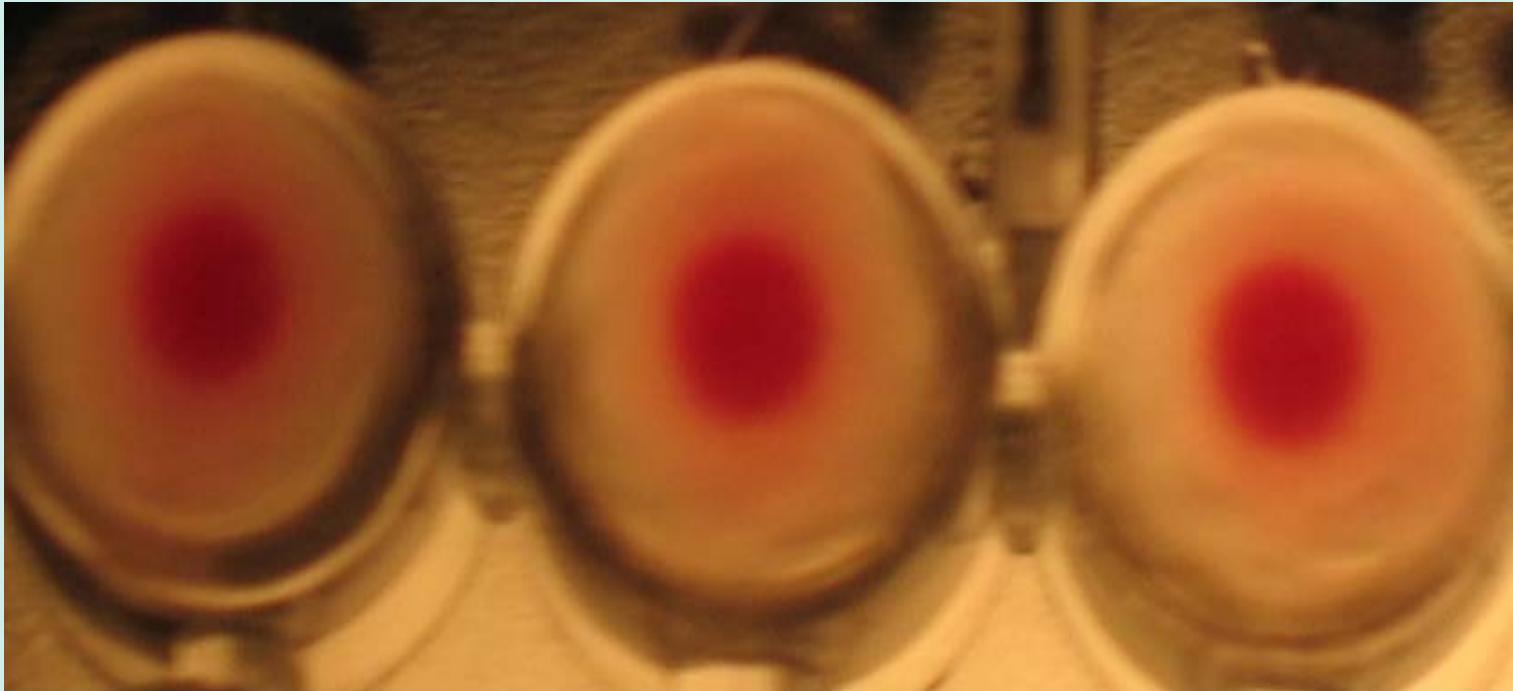


C **c** **D** **E** **e** **ctl**

human/humain

E000 206 10.95

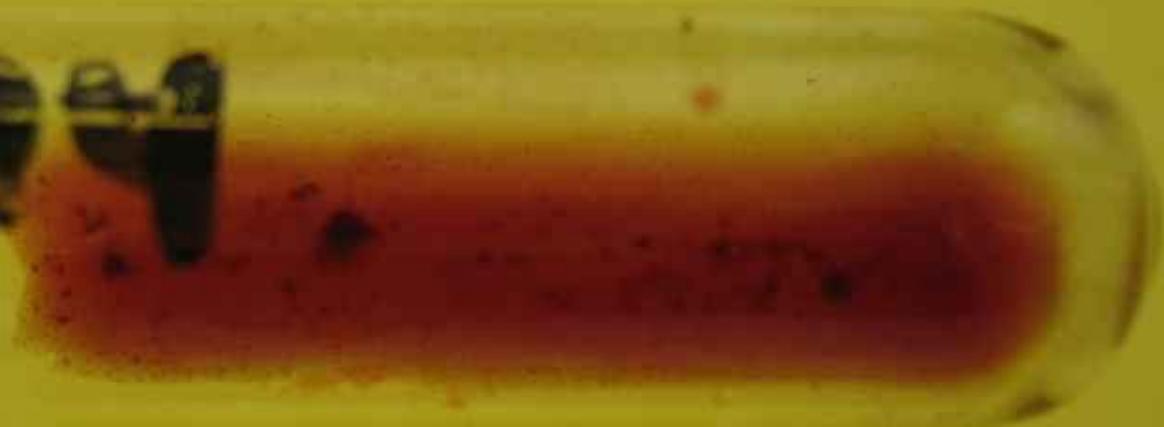
627-9624



microwell plate
antisera dilution 1:5

negative reaction

৭০০০



Weak D-TYPE

Worksheet und Auswertetabelle / Worksheet and Evaluation diagram

Reaktions-Nr. / Reaction No.	1	2	3	4	5	6	7	8
PCR-Produkt (Größe in bp) PCR product (size in bp)	150	126	165	101	130	112	198	153
weak D Allele / weak D alleles								
weak D type 1	+	-	-	-	-	-	-	-
weak D type 2	-	+	-	-	-	-	-	-
weak D type 3	-	-	+	-	-	-	-	-
weak D type 4.0, 4.1	-	-	-	+	-	-	-	-
weak D type 4.2, DAR	-	-	-	+	+	-	-	-
weak D type 5	-	-	-	-	-	+	-	-
weak D type 11 (haplotyp cDe)	-	-	-	-	-	-	+	-
RHD(M295I) (haplotyp CD ₄ e)	-	-	-	-	-	-	+	-
weak D type 15	-	-	-	-	-	-	-	+
RHD pos. oder / or RHD neg.	-	-	-	-	-	-	-	-

Ergebnis Result	1	2	3	4	5	6	7	8	Genotyp Genotype

Proben-ID / Sample-ID: 0627 9624

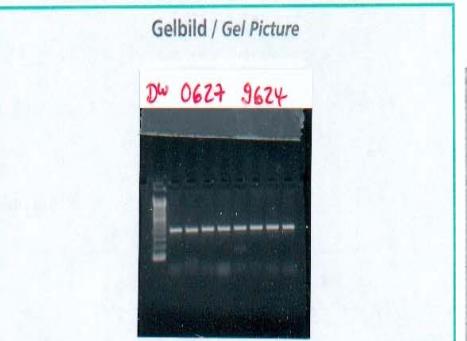
Name: [Signature]

Geb.-Datum / Birthdate:

Ergebnis / Result: \Rightarrow D. partial

Datum / Date: 25.10.04

Unterschrift / Signature: [Signature]



Partial D-TYPE

Worksheet und Auswertetabelle / Worksheet and Evaluation diagram

Reaktions-Nr. / Reaction No.	1	2	3	4	5	6	7	8	9	10	11
PCR-Produkt (Größe in bp) PCR product (size in bp)	146	118	135	132	132	120	166	117	140	107	113
RHD Exons	D ₂	D ₃	D ₄	D ₅	D ₆	D ₇	D ₂	D ₇	D ₈	D ₆	D ₉
Phänotypen / Phenotypes	Standard RHD										
D	+	+	+	+	+	+	-	-	-	-	-
RHD Varianten / RHD Variants											
D cat. IIIa, IIIc, III type IV n.t.	+	-	+	+	+	+	-	-	-	-	-
D cat. IIb n.t.	-	+	+	+	+	+	-	-	-	-	-
D cat. IVa n.t.	+	-	+	+	+	-	-	-	-	-	-
D cat. IVb, IVb(I), IV type IV n.t.	+	+	+	+	+	-	-	-	-	-	-
D cat. IV type III	+	+	+	+	-	-	-	-	-	-	-
D cat. Va, Va-like, Va-associated	+	+	+	-	+	+	-	-	-	-	-
D cat. VI type I	+	+	-	-	+	+	-	-	-	-	-
D cat. VI type II	+	+	-	-	-	+	-	-	-	-	-
D cat. VI type III	+	-	-	-	-	+	-	-	-	-	-
D cat. VI type IV n.t.	-	-	-	-	+	+	-	-	-	-	-
D cat. VII	+	+	+	+	+	+	-	-	-	-	-
DAR	+	+	+	+	+	+	-	-	-	-	-
DAU	+	+	+	+	+	+	-	-	+	-	-
DBT	+	+	+	-	-	-	-	-	-	-	-
DFR	+	+	-	+	+	+	-	-	-	-	-
DHMi	+	+	+	+	+	+	-	-	-	+	-
DHMi' n.t.	+	-	-	-	+	+	-	-	-	-	-
DNB	+	+	+	+	+	+	-	+	-	-	-
ROHar (Rh33)	-	-	-	+	-	-	-	-	-	-	-
D cat. II, DCS, DFW, DHR, DIM, DNU n.t.	+	+	+	+	+	+	-	-	-	-	-
Rh D ^a (RHD(K409K))	+	+	+	+	+	+	-	-	-	-	+

n.t. = not tested currently

Ergebnis Result	1	2	3	4	5	6	7	8	9	10	11	Phänotyp Phenotype	Genotyp Genotype

Proben-ID / Sample-ID: 0627 9624

Name: [Signature]

Geb.-Datum / Birthdate:

Ergebnis / Result: D. VI

Datum / Date: 26.10.04

Unterschrift / Signature: [Signature]



Weak D-TYPE

Worksheet und Auswertetabelle / Worksheet and Evaluation diagram

Reaktions-Nr. / Reaction No.
PCR-Produkt (Größe in bp) PCR product (size in bp)
weak D type 1
weak D type 2
weak D type 3
weak D type 4.0, 4.1
weak D type 4.2, DAR
weak D type 5
weak D type 11 (haplotyp)
RHD(M295I) (haplotyp)
weak D type 15
RHD pos. oder / or RHD

Final result: D category VI type II

Rhesus prophylaxis required

Transfusion of D negative red blood cells is required

Ergebnis Result	1	2	3	4	5	6	7	8	Genotyp Genotype

Proben-ID / Sample-ID: 0627 9624

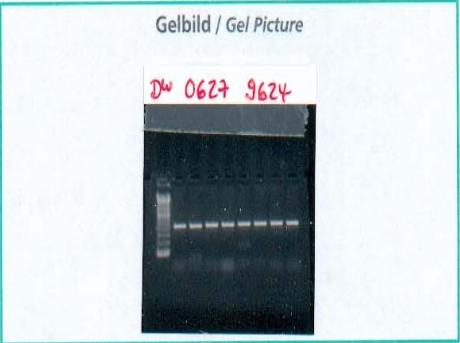
Name:

Geb.-Datum / Birthdate:

Ergebnis / Result: \Rightarrow D. partial

Datum / Date: 25.10.04

Unterschrift / Signature: 



Proben-ID / Sample-ID: 0627 9624

Name:

Geb.-Datum / Birthdate:

Ergebnis / Result: D. VI

Datum / Date: 26.10.04

Unterschrift / Signature: 



Partial D-TYPE

Worksheet und Auswertetabelle / Worksheet and Evaluation diagram

Reaktions-Nr. / Reaction No.	1	2	3	4	5	6	7	8	9	10	11
PCR-Produkt (Größe in bp) PCR product (size in bp)	146	118	135	132	132	120	166	117	140	107	113
α	-	-	-	-	-	-	-	-	-	-	-

Contents of our BAGene SSP kits

- PCR plates or strips with pre aliquoted, dried and colored reaction mixes containing allele specific primers, internal control primers (specific for the HGH gene) and nucleotides.
- 10 x PCR buffer
- PCR strip caps
- Worksheets and Evaluation Diagrams
- Instructions for Use

Evaluation of Results - RHD-RHCE genotyping



BAGene RH-TYPE

Worksheet und Auswertetabelle / Worksheet and Evaluation diagram

Reaktions-Nr. / Reaction No.	1	2	3	4	5	6	7	8	9	10	11	12	13
PCR-Produkt/e / PCR product/s (Größe in bp/size in bp)	224 123 154	390 248	140	113	113	198	143	215	162	145	157	155	181
Reaktionsmuster / Reaction pattern	D _{4/4} RHD(W16X) RHD Ψ	D ₇	D ₄ RHD Ψ neg. (K409K) neg.	D _{el} RHD (K409K) neg.	D _d RHD (M295) neg.	D _{el} RHD (IV53+1G>A)	D _d RHD (IV53+1G>A)	Cde ^s	C ^c C ^w	c	E	e	C ^w
Phänotyp / Phenotype	Genotyp / Genotype	Standard RHD/RHCE Allele/s											
D-positive	+ +	+	+ +	-	-	-	-	-	-	-	-	-	
D-negative	- -	-	- -	-	-	-	-	-	-	-	-	-	
C	Größe der internen PCR-Kontrolle in Reaktion Nr. 2: 659 bp, in allen anderen Reaktionen: 434 bp (GHG)												
C ^w	Size of internal PCR control in reaction no. 2: 659 bp, in all the other reactions: 434 bp (GHG)												
c													
E													
e													
RHD-Varianten / RHD variants													
D-positive	RHD / RHD(K409K)	+ +	+	+ +	+ +	-	-	-	-	-	-	-	-
D ₄	RHD(K409K)	+ +	+	+ -	+ +	-	-	-	-	-	-	-	-
D-positive	RHD / RHD(M295)	+ +	+	+ +	+ -	+ +	-	-	-	-	-	-	-
D _d	RHD(M295)	+ +	+	+ +	+ -	+ +	-	-	-	-	-	-	-
D-positive	RHD / RHD(IV53+1G>A)	+ +	+	+ +	+ -	-	-	+ +	-	-	-	-	-
D ₄	RHD / RHD(IV53+1G>A)	+ +	+	+ +	+ -	-	-	+ +	-	-	-	-	-
D-positive	RHD / RHD Ψ	+ +	+ +	+ +	+ -	-	-	-	-	-	-	-	-
D-negative	RHD Ψ	+ +	+ +	+ -	-	-	-	-	-	-	-	-	-
D-negative	RHD-CE(8-9)-D	+ +	-	+ +	-	-	-	-	-	-	-	-	-
D-negative	RHD(W16X)	+ +	+ +	+ +	-	-	-	-	-	-	-	-	-
D-positive	RHD / Cde ^s	+ +	+	+ +	+ -	-	-	-	-	-	-	-	-
D-negative	Cde ^s / Cde ^s ; Cde ^s /d	-	+ +	-	+ +	-	-	-	-	-	-	-	-
Beispiele / Examples													
ccdee	-	-	-	-	-	-	-	-	-	-	-	-	-
Ccdee	-	-	-	-	-	-	-	-	-	-	-	-	-
CcDEe	+ +	+	+ +	-	-	-	-	-	-	-	-	-	-
CCDE.ee	+ +	+	+ +	-	-	-	-	-	-	-	-	-	-
CC ^w D.Ee	+ +	+	+ +	-	-	-	-	-	-	-	-	-	-
D cat. VI	+ +	-	+ +	-	-	-	-	-	-	-	-	-	-
D cat. IV type III	-	-	+ +	+ +	-	-	-	-	-	-	-	-	-
DFR	+ +	+	-	+ +	-	-	-	-	-	-	-	-	-

Fehlende Banden in den Reaktionen Nr. 1 - 4 können auf RHD psi oder auf Partial D-Gene hinweisen.
In Gegenwart von Cde^s kommt es zum Ausfall der Reaktion Nr. 9.
Missing bands in reaction no. 1 - 4 may either indicate RHD psi or partial D genes.
In presence of Cde^s reaction no. 9 is missing.

Ergebnis / Result	1	2	3	4	5	6	7	8	9	10	11	12	13	Phänotyp / Phenotype	Genotyp / Genotype

Proben-ID / Sample-ID:	Gelbild / Gel Picture											
Probenname / Name:												
Geb.-Datum / Born:												
Ergebnis / Result:												
Datum / Date:												
Unterschrift / Signature:												

VER 05/2006

Kurzanleitung / Short Instructions

1. Die gewünschte Anzahl **BAGene**-Platten/-Streifen aus dem Gefrierschrank (-20...-80°C) nehmen und den 10 x PCR-Puffer bei Raumtemperatur auftauen.

Remove the required number of **BAGene**-plates/-strips from -20...-80°C and thaw the 10 x PCR-buffer.

2. Der erste Reaktionsmix ist markiert (PCR-Streifen: schwarzer Strich um das erste Reaktionsgefäß, PCR-Platten: schwarzer Punkt oberhalb des ersten Reaktionsgefäßes).

The first reaction mix is marked (PCR-strips: black line around the first reaction tube, PCR-plates: black dot above the first reaction tube).

3. Den Mastermix, bestehend aus 10 x PCR-Puffer, DNA-Lösung, Taq-Polymerase und Aqua dest., zusammenpipettieren und gründlich vortoxen. Die verschiedenen **BAGene** DNA-SSP Kits werden mit dem gleichen Mastermix angesetzt und sind daher miteinander kombinierbar.

Pipet the mastermix, consisting of 10 x PCR-buffer, DNA-solution, Taq-Polymerase and aqua dest. and mix well.

The different **BAGene** DNA-SSP Kits do all work with the same mastermix and can therefore be combined.

Zusammensetzung des Mastermixes in Abhängigkeit von der Anzahl der Reaktionsmixe

Composition of the Mastermix depending on the number of reaction mixes

No. of mixes	DNA-sol. (50-100 ng/µl)	Aqua dest.	10 x PCR-buffer	Taq-Polymerase (5 µl)	total volume app.	
1	1	8	1	0,08	10	µl
2	2	16	2	0,2	20	µl
6*	7	50	7	0,5	65	µl
7	9	70	9	0,7	90	µl
8	10	80	10	0,8	100	µl
9	11	88	11	0,9	110	µl
10	12	96	12	1,0	120	µl
11	13	104	13	1,0	130	µl
12	14	112	14	1,1	140	µl
13	16	128	16	1,3	160	µl
14	17	136	17	1,4	170	µl
15	18	144	18	1,4	180	µl
16	19	152	19	1,5	190	µl

Bei abweichenden DNA-Konzentrationen sind die Mengen von DNA und Wasser entsprechend zu variieren.
For different DNA concentrations, the quantities of DNA and water must be varied accordingly.

* Mastermix für 6 Reaktionsmixe wird wegen des geringen Volumens Taq-Polymerase als Mindestansatz empfohlen.

Minimum preparation of mastermix for 6 reaction mixes is recommended due to the small volume of Taq-Polymerase.

4. Nach dem Vortexen werden von diesem Gemisch umgehend je 10 µl zu den angetrockneten Reaktionsmixen pipettiert.
After vortexing add 10 µl of this mixture immediately to the dried reaction mixes.

5. Die Reaktionsgefäße werden mit den dafür vorgesehenen Deckeln **dicht** verschlossen. Die Platten/Streifen werden leicht bewegt, um das Pellet auf dem Gefäßboden etwas anzulösen. Die gesamte Reaktionslösung soll sich im Gefäßboden befinden.
Tightly close the tubes with the respective strip caps. Slightly move the plates/strips to dissolve the pellet at the bottom of the tube. All PCR-solution should settle on the bottom of the tube.

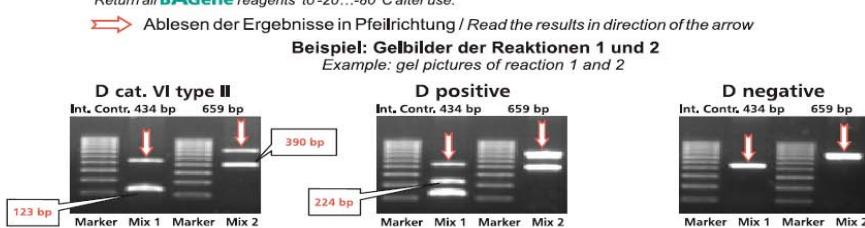
6. Nach der PCR und Auftrennung der Amplikate im Gel erfolgt die Auswertung.
After PCR and separation of the amplicons in the gel, the evaluation can be carried out.

7. Alle **BAGene** Reagenzien sind nach Gebrauch wieder bei -20...-80°C zu lagern.
Return all **BAGene** reagents to -20...-80°C after use.

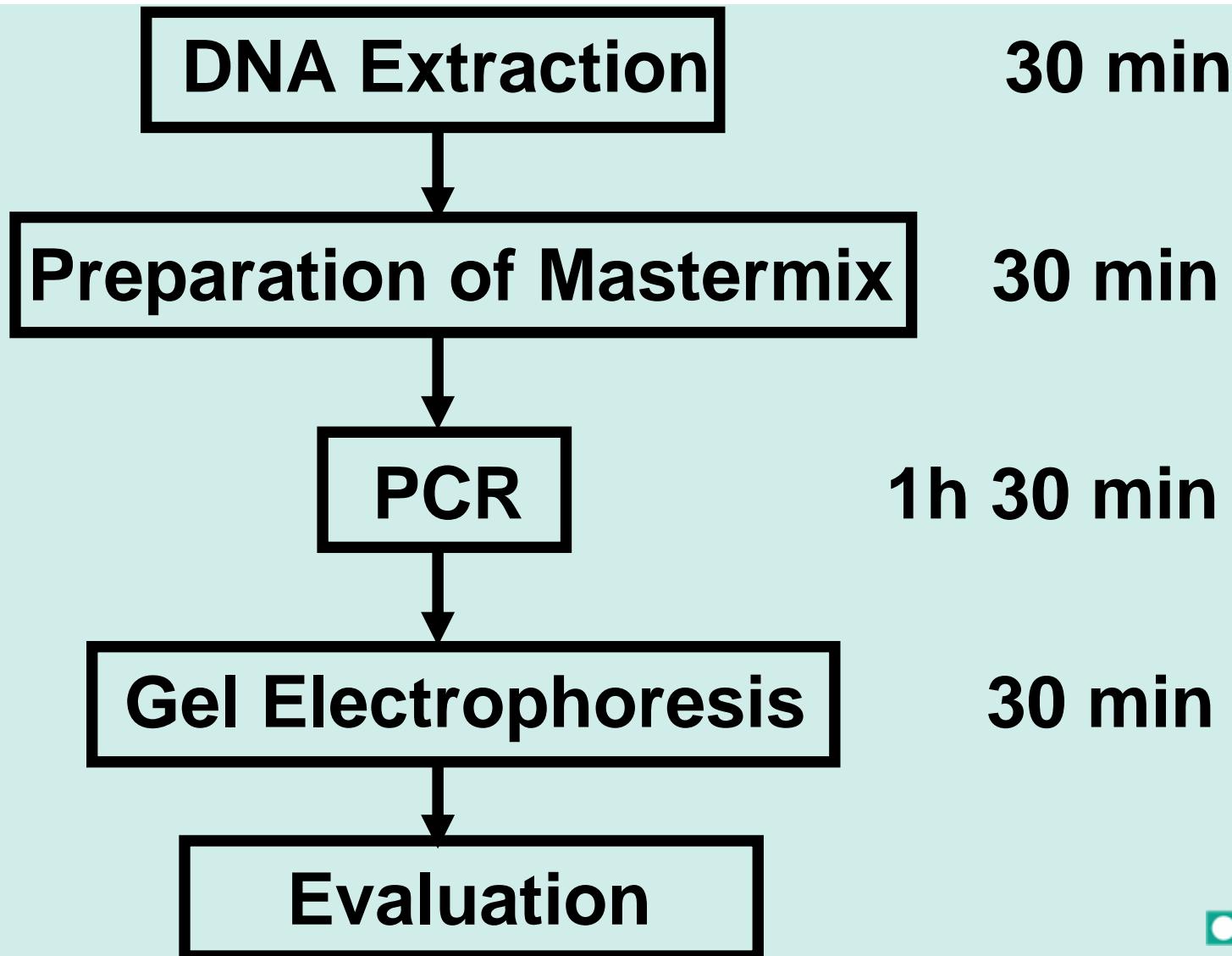
→ Ablesen der Ergebnisse in Pfeilrichtung / Read the results in direction of the arrow

Beispiel: Gelbilder der Reaktionen 1 und 2

Example: gel pictures of reaction 1 and 2

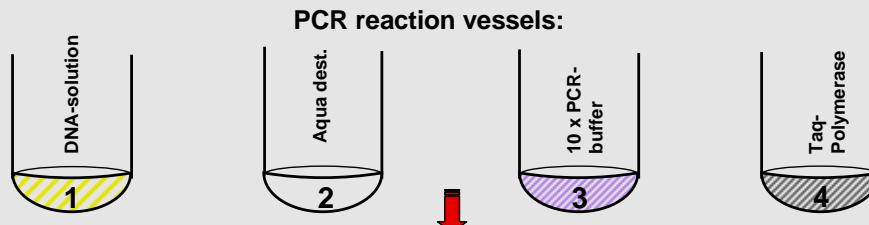


Test Procedure - Workflow



Preparation of the PCR reaction mixes

BAGene DNA-SSP Kits Pipetting Chart

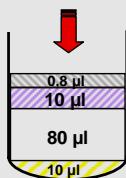


Preparation of the mastermix depending on the number of reaction mixes

	No. of mixes	DNA-solution (50-100 ng/ μ l)	Aqua dest.	10 x PCR-buffer	Taq-Polymerase (5 U/ μ l)	total volume, app.	
D Zygoty-TYPE, MNS-TYPE preparation for 6 mixes	6	7	50	7	0,5	65	μ l
RHD-TYPE Asia - 7 mixes	7	9	70	9	0,7	90	μ l
ABO-, KKD-, or Weak D-TYPE - 8 mixes	8	10	80	10	0,8	100	μ l
HPA-TYPE - 12 mixes	12	14	112	14	1,1	140	μ l
RH-TYPE - 13 mixes	13	16	128	16	1,3	160	μ l
ABO-TYPE variant or Partial D-TYPE - 16 mixes	16	19	152	19	1,5	190	μ l

reaction mix

e.g. ABO-TYPE, KKD-TYPE or Weak D-TYPE

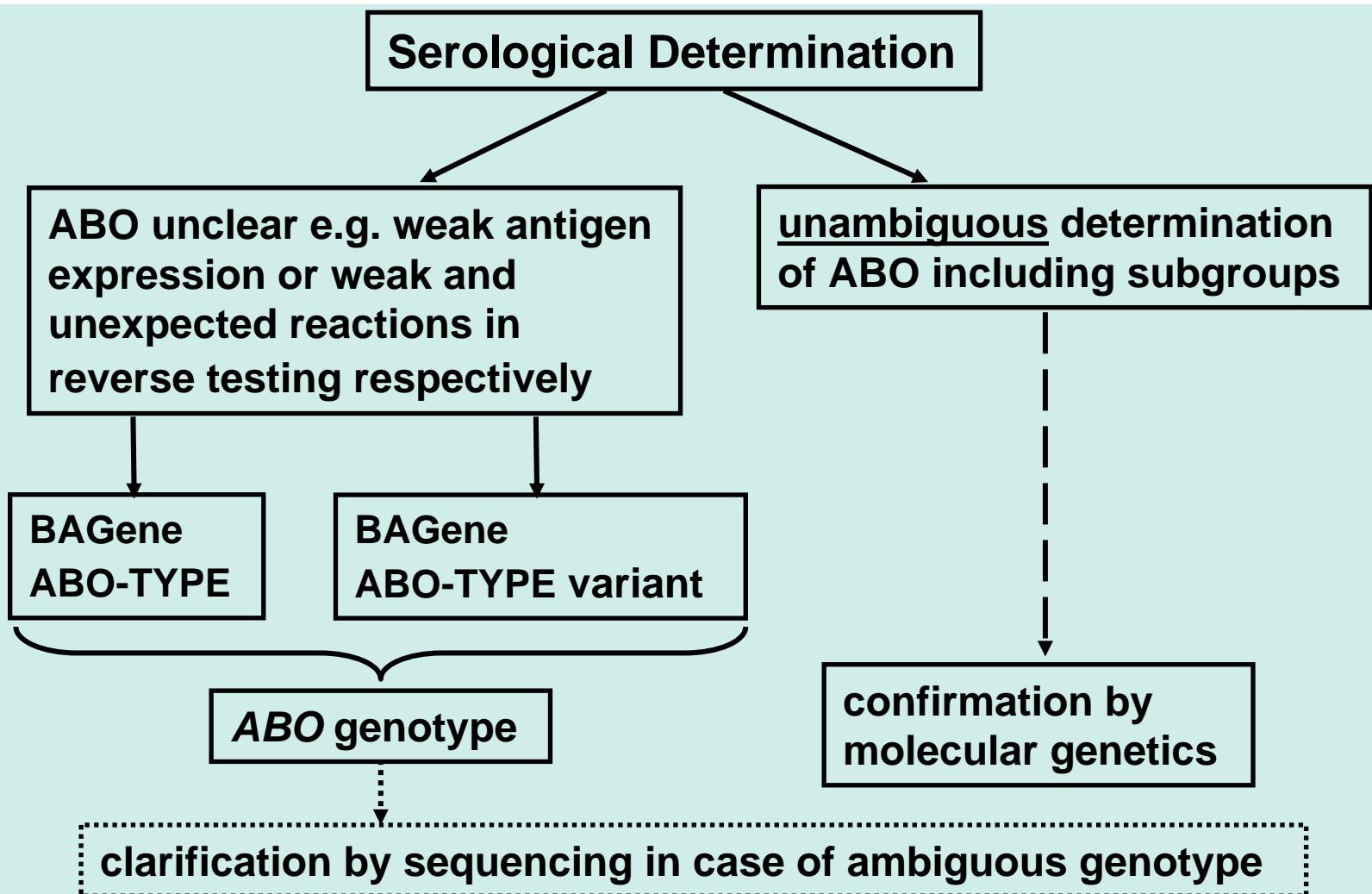


mix well (vortex)

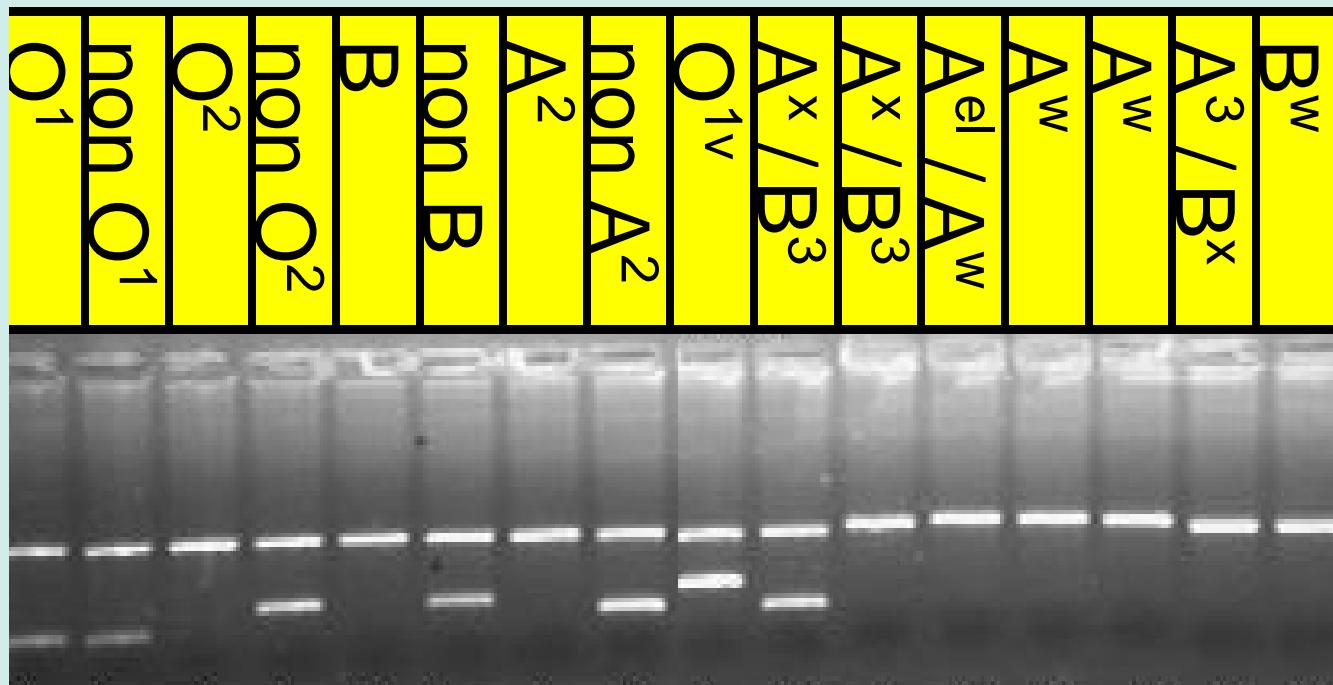
add 10 μ l to each well of the PCR-strips/-plates

Investigation Strategy

♦ ABO blood group typing



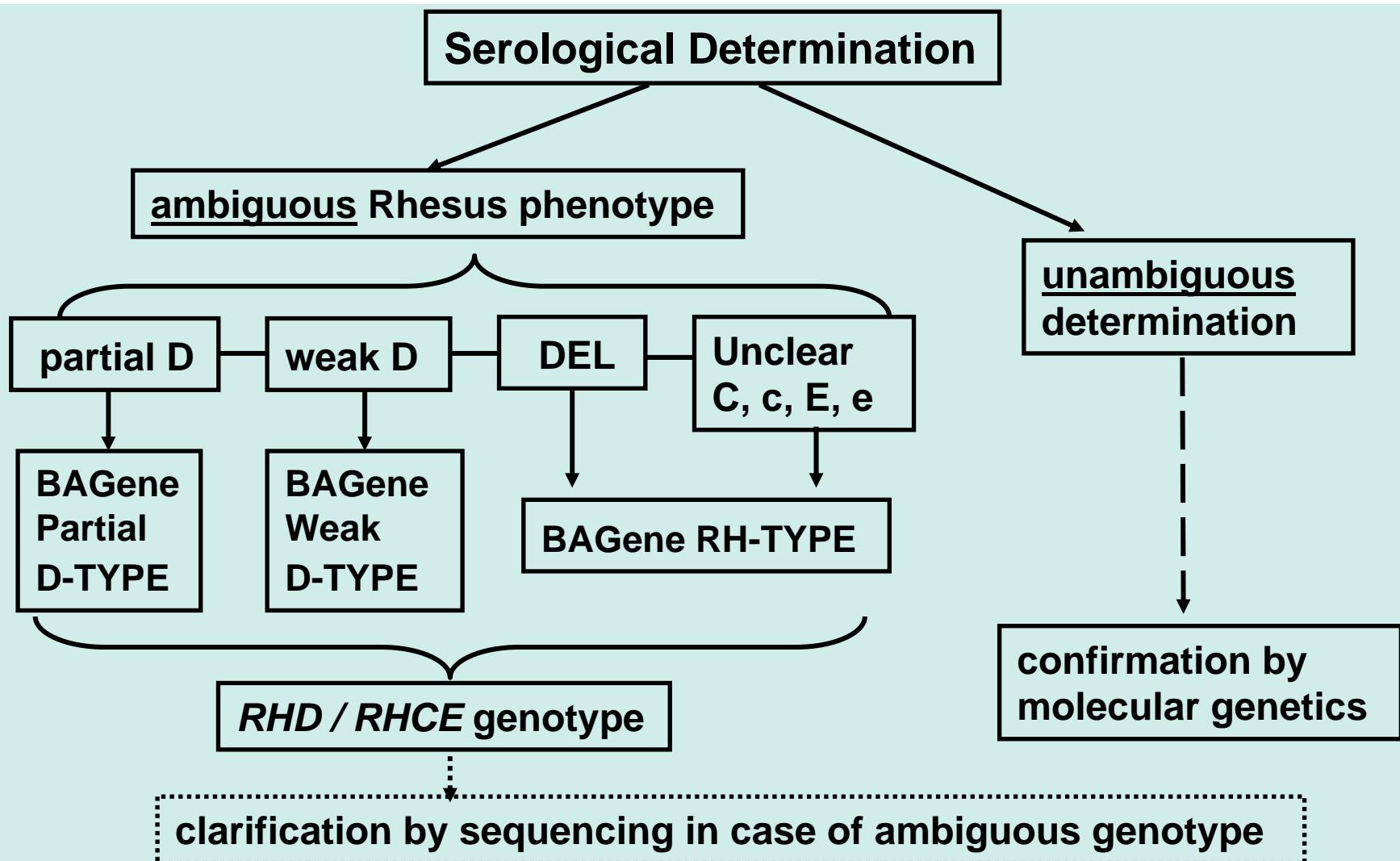
ABO Genotyping



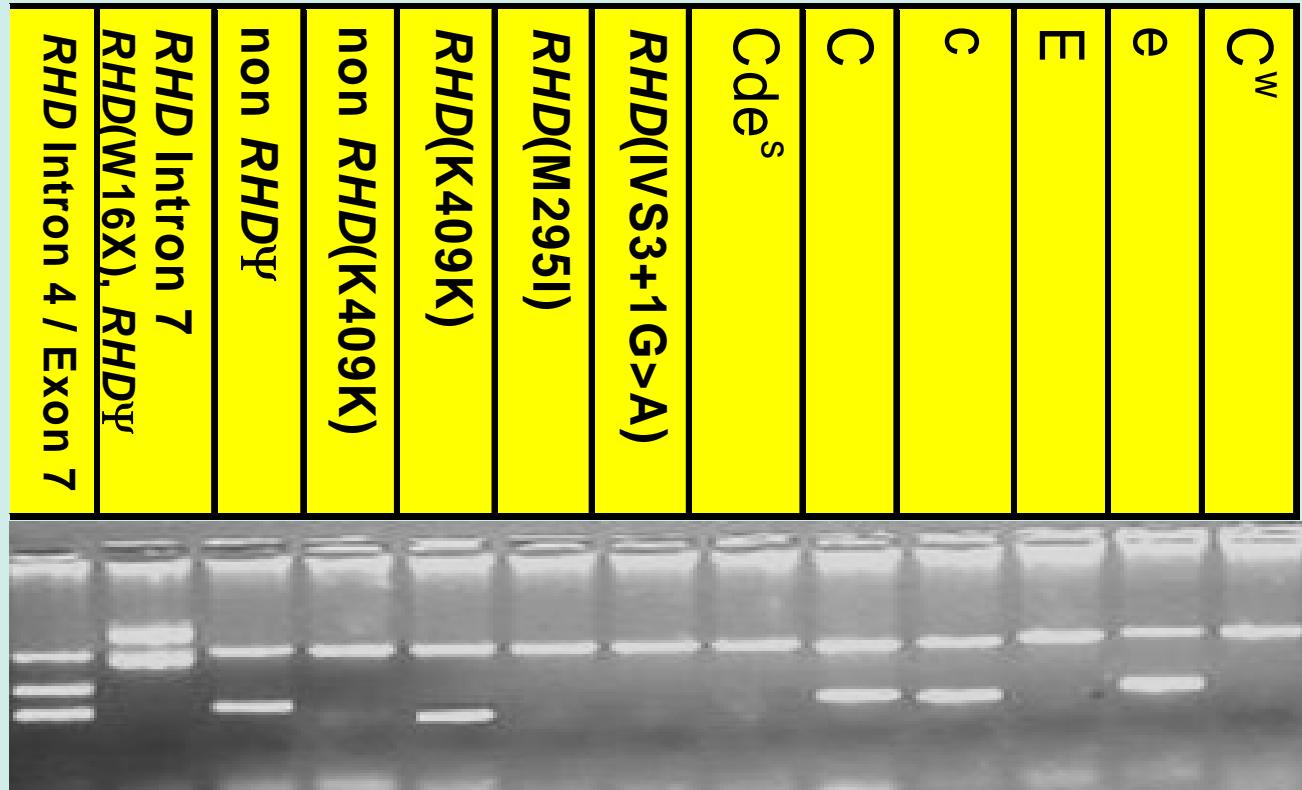
BAGene ABO-TYPE variant genotype $O^{1v}A^x$

Investigation Strategy

♦ Rhesus typing

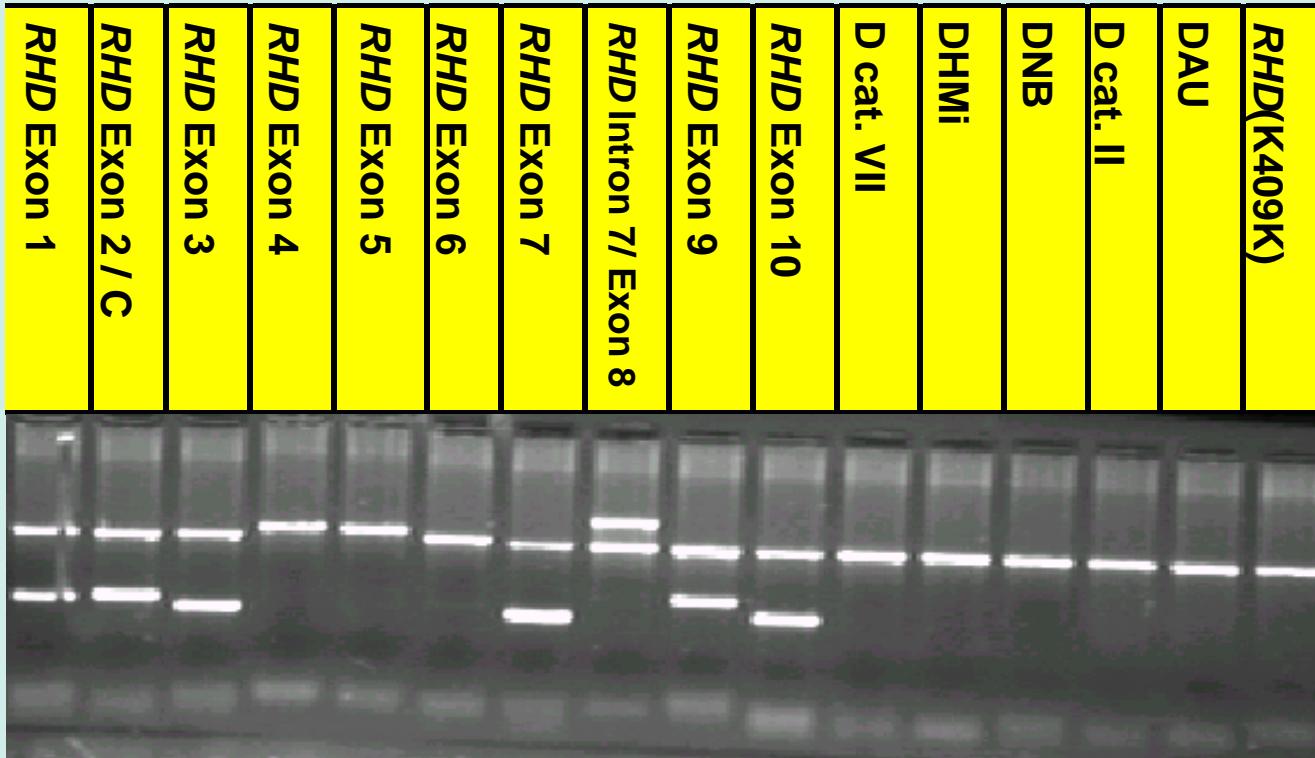


RHD, RHCE Genotyping



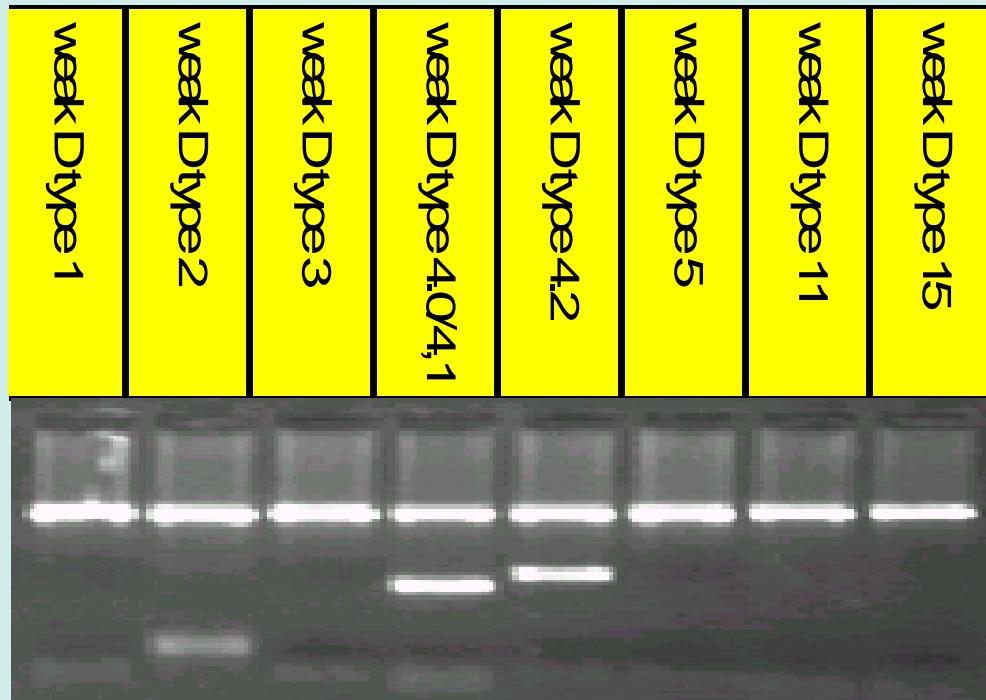
BAGene RH-TYPE genotype *RHD(K409K) Ccee*

Genotyping partial D



BAGene Partial D-TYPE genotype *D cat. VI type II*

Genotyping weak D



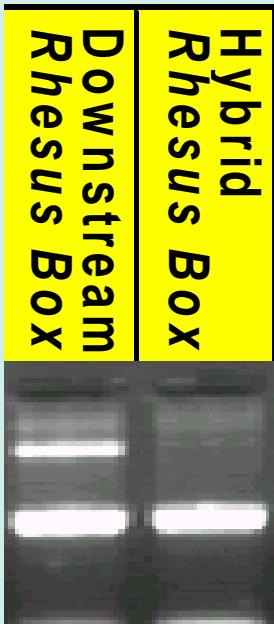
BAGene Weak D-TYPE genotype *weak D type 4.2*

RHD Genotyping „Asia“

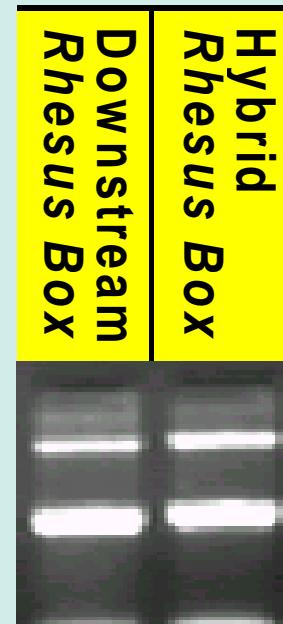


BAGene RHD-TYPE Asia genotype RHD positive / weak D type 17

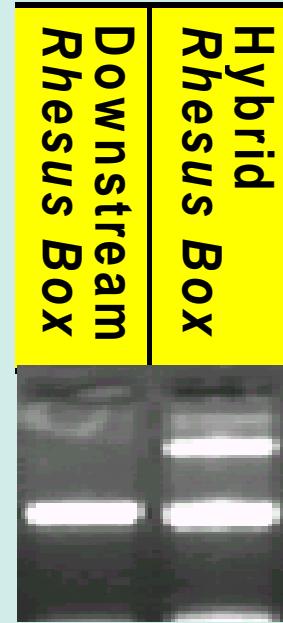
RHD Zygosity



DD



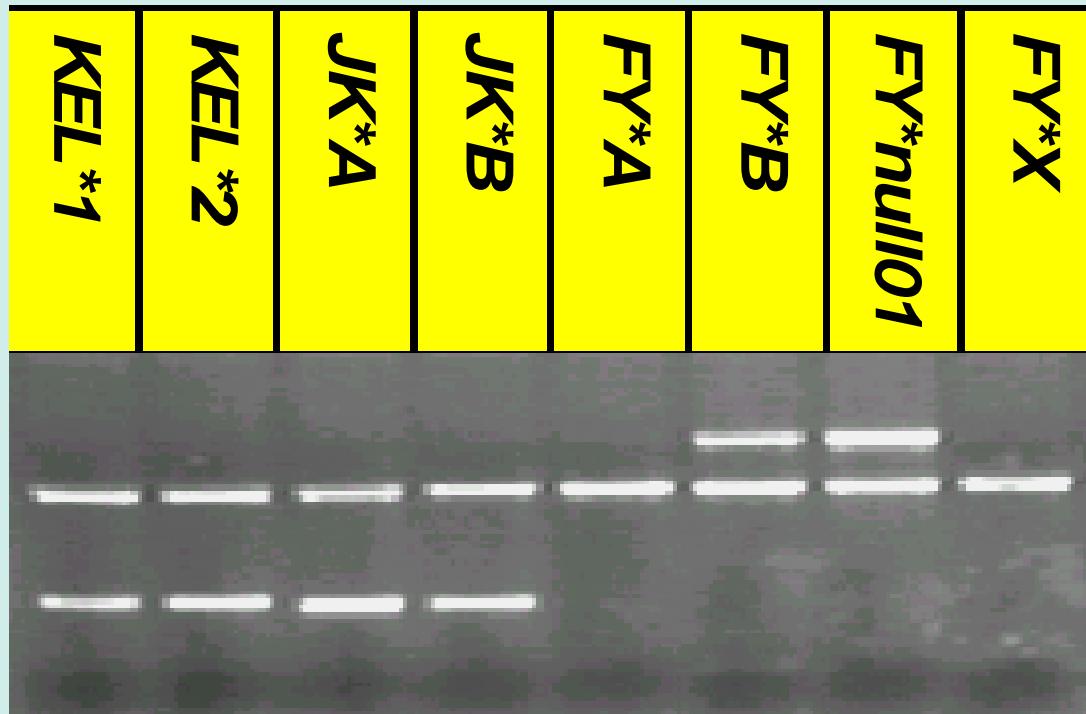
Dd



dd

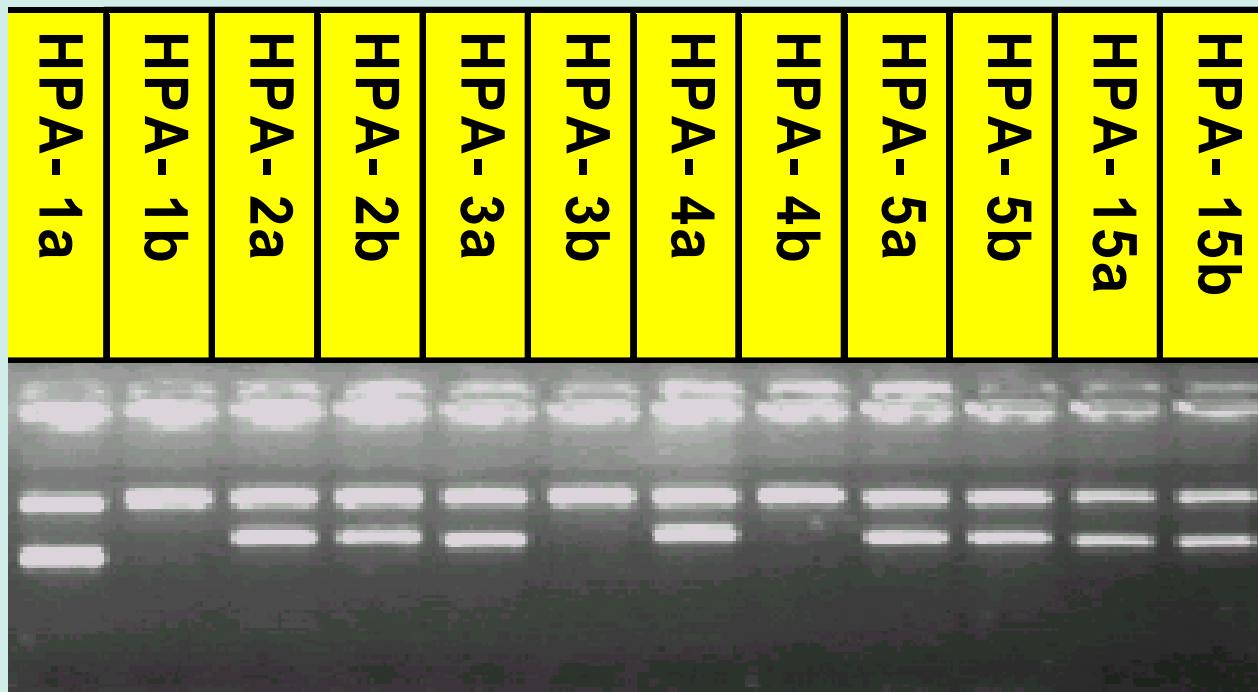
BAGene D Zygosity-TYPE

Genotyping *KEL*, *JK*, *FY*



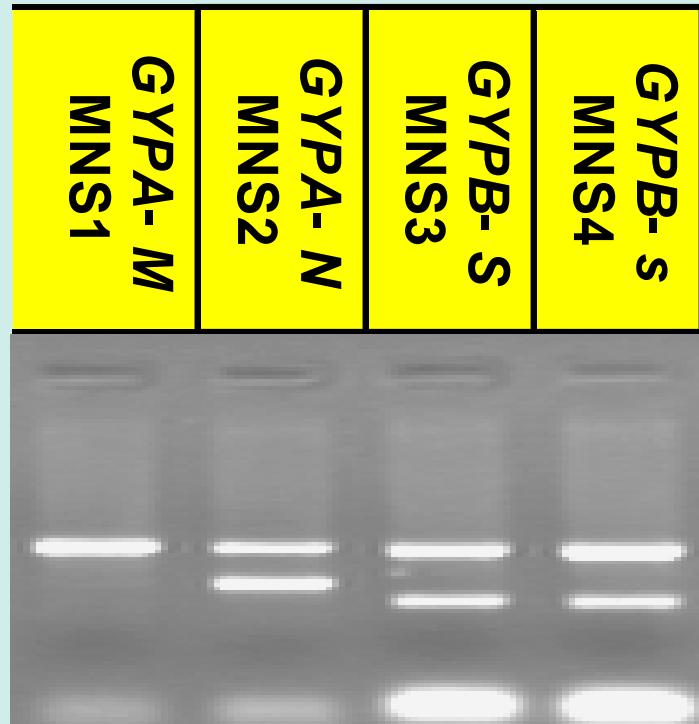
BAGene KKD-TYPE
genotype *KEL**1/*KEL**2; *JK**A/*JK**B; *FY**B/*FY**null01

Genotyping *HPA*



BAGene HPA-TYPE
genotype *HPA-1a/a; 2a/b; 3a/a; 4a/a; 5a/b; 15 a/b*

Genotyping MNS



BAGene MNS-TYPE
genotype *NSs*

Recently published, presented soon

► Molecular genetic blood group typing by the use of PCR-SSP technique

M Prager, BAG Health Care, Lich, Germany

presented at the FDA Workshop on Molecular Methods in Immunohematology

Bethesda 25 – 26 Sept 2006

published in Transfusion 2007;47:54S-59S.

► ABO genotyping for diagnosis of unusual ABO blood groups:

A Comparative Study in German Blood Donor Centers

M Prager, EA Scharberg, FF Wagner, J Burkhardt, A Seltsam

Presentation at 40. Annual Meeting of the German Society for Transfusion Medicine
and Immunohematology (DGTI) September 18-21, 2007